

Pan-cancer: <i>BRAF</i> , <i>NTRK1</i> , <i>NTRK2</i> , <i>NTRK3</i> , <i>RET</i> , MSI, TMB											
Genes with biomarkers of clinical significance*											Genes with biomarkers of potential clinical significance†
	Breast	<i>BRCA1</i>	<i>BRCA2</i>	<i>ERBB2</i>	<i>ESR1</i>	<i>PALB2</i>	<i>PIK3CA</i>				180
	Colorectal	<i>ERBB2</i>	<i>KRAS</i>	<i>NRAS</i>							166
	Bone	<i>EGFR</i>	<i>ERG</i>	<i>ETV1</i>	<i>ETV4</i>	<i>EWSR1</i>	<i>FEV</i>	<i>FLI1</i>	<i>FUS</i>	<i>H3F3A</i>	140
	Lung	<i>ALK</i>	<i>EGFR</i>	<i>ERBB2</i>	<i>KRAS</i>	<i>MET</i>	<i>NUTM1</i>	<i>ROS1</i>			223
	Melanoma	<i>KIT</i>	<i>NRAS</i>	<i>ROS1</i>							172
	Ovarian	<i>BRCA1</i>	<i>BRCA2</i>	<i>FOXL2</i>							149
	CNS‡	<i>APC</i>	<i>ATRX</i>	<i>CDKN2A</i>	<i>CDKN2B</i>	<i>EGFR</i>	<i>H3F3A</i>	<i>HIST1H3B</i>	<i>HIST1H3C</i>	<i>IDH1</i>	140
	Prostate	<i>AR</i>	<i>ATM</i>	<i>BARD1</i>	<i>BRCA1</i>	<i>BRCA2</i>	<i>BRIP1</i>	<i>CDK12</i>	<i>CHEK1</i>	<i>CHEK2</i>	151
	Thyroid	<i>HRAS</i>	<i>KRAS</i>	<i>NRAS</i>	<i>TERT</i>						165
	Uterine & cervical	<i>BRCA2</i>	<i>EPC1</i>	<i>ERBB2</i>	<i>ESR1</i>	<i>FOXO1</i>	<i>GREB1</i>	<i>JAZF1</i>	<i>NCOA2</i>	<i>NCOA3</i>	138
	Other solid tumors	<i>ALK</i>	<i>APC</i>	<i>ARID1A</i>	<i>ASPSR1</i>	<i>ATF1</i>	<i>ATIC</i>	<i>BAP1</i>	<i>BCOR</i>	<i>BRCA1</i>	152
		<i>CARS</i>	<i>CCNB3</i>	<i>CDK4</i>	<i>CDKN2A</i>	<i>CIC</i>	<i>CITED2</i>	<i>CLTC</i>	<i>COL1A1</i>	<i>COL6A3</i>	
		<i>CREB3L2</i>	<i>CSF1</i>	<i>CTNNB1</i>	<i>DDIT3</i>	<i>DDX3X</i>	<i>DNAJB1</i>	<i>DUX4</i>	<i>EED</i>	<i>EGFR</i>	
		<i>ETV1</i>	<i>ETV4</i>	<i>ETV6</i>	<i>EWSR1</i>	<i>FEV</i>	<i>FGFR2</i>	<i>FGFR3</i>	<i>FLI1</i>	<i>FOXL2</i>	
		<i>FUS</i>	<i>GLI1</i>	<i>HEY1</i>	<i>HGF</i>	<i>HMGA2</i>	<i>IDH1</i>	<i>KRAS</i>	<i>LEUTX</i>	<i>MAML3</i>	
		<i>MYOD1</i>	<i>NAB2</i>	<i>NCOA2</i>	<i>NF1</i>	<i>NFATC2</i>	<i>NFIB</i>	<i>NR4A3</i>	<i>NRAS</i>	<i>NUTM1</i>	
		<i>PALB2</i>	<i>PATZ1</i>	<i>PAX3</i>	<i>PAX7</i>	<i>PDGFB</i>	<i>PDGFRA</i>	<i>PRKACA</i>	<i>PRKD1</i>	<i>RANBP2</i>	
		<i>SDHB</i>	<i>SDHC</i>	<i>SDHD</i>	<i>SMARCB1</i>	<i>SS18</i>	<i>SSX1</i>	<i>SSX2</i>	<i>SSX4</i>	<i>STAT6</i>	
		<i>TCF12</i>	<i>TERT</i>	<i>TFE3</i>	<i>TFEB</i>	<i>TFG</i>	<i>TP53</i>	<i>TPM3</i>	<i>TPM4</i>	<i>TRAF7</i>	
		<i>WT1</i>	<i>WWTR1</i>	<i>YAP1</i>	<i>YWHAE</i>	<i>ZC3H7B</i>					

Figure 3: Subset of genomic tumor profiling biomarkers for multiple cancer types—TruSight Oncology 500 and TruSight Oncology 500 High-Throughput include key guideline biomarkers, emerging biomarkers, and pan-cancer biomarkers such as *BRAF*, *NTRK1*, *NTRK2*, *NTRK3*, *RET*, MSI, and TMB. Content analysis provided by Velsera based on software Knowledge Base v8.5 (February 2023).

* Genes with biomarkers of clinical significance linked to current drug labels or guidelines.

† Genes with biomarkers of potential clinical significance based on presence in clinical trials.

‡ CNS, central nervous system.

Table 2: DNA content included in TruSight Oncology 500 and TruSight Oncology 500 High-Throughput

ABL1	BCR	CHEK1	EPHA7	FGF23	GSK3B	IDH2	MAP3K1	NF2	PIK3CA	RAD51D	SMAD4	TGFBR2
ABL2	BIRC3	CHEK2	EPHB1	FGF3	H3F3A	IFNGR1	MAP3K13	NFE2L2	PIK3CB	RAD52	SMARCA4	TMEM127
ACVR1	BLM	CIC	ERBB2	FGF4	H3F3B	INHBA	MAP3K14	NFKBIA	PIK3CD	RAD54L	SMARCB1	TMPRSS2
ACVR1B	BMPR1A	CREBBP	ERBB3	FGF5	H3F3C	INPP4A	MAP3K4	NKX2-1	PIK3CG	RAF1	SMARCD1	TNFAIP3
AKT1	BRAF	CRKL	ERBB4	FGF6	HGF	INPP4B	MAPK1	NKX3-1	PIK3R1	RANBP2	SMC1A	TNFRSF14
AKT2	BRCA1 ^a	CRLF2	ERCC1	FGF7	HIST1H1C	INSR	MAPK3	NOTCH1	PIK3R2	RARA	SMC3	TOP1
AKT3	BRCA2 ^a	CSF1R	ERCC2	FGFR1	HIST1H2BD	IRF2	MAX	NOTCH2	PIK3R3	RASA1	SMO	TOP2A
ALK	BRD4	CSF3R	ERCC3	FGFR2	HIST1H3A	IRF4	MCL1	NOTCH3	PIM1	RB1	SNCAIP	TP53
ALOX12B	BRIP1	CSNK1A1	ERCC4	FGFR3	HIST1H3B	IRS1	MDC1	NOTCH4	PLCG2	RBM10	SOCS1	TP63
ANKRD11	BTG1	CTCF	ERCC5	FGFR4	HIST1H3C	IRS2	MDM2	NPM1	PLK2	RECQL4	SOX10	TRAF2
ANKRD26	BTB	CTLA4	ERG	FH	HIST1H3D	JAK1	MDM4	NRAS	PMAIP1	REL	SOX17	TRAF7
APC	C11orf30	CTNNA1	ERF1	FLCN	HIST1H3E	JAK2	MED12	NRG1	PMS1	RET	SOX2	TSC1
AR	CALR	CTNNB1	ESR1	FLI1	HIST1H3F	JAK3	MEF2B	NSD1	PMS2	RFXD2	SOX9	TSC2
ARAF	CARD11	CUL3	ETS1	FLT1	HIST1H3G	JUN	MEN1	NTRK1	PNRC1	RHEB	SPEN	TSHR
ARFRP1	CASP8	CUX1	ETV1	FLT3	HIST1H3H	KAT6A	MET	NTRK2	POLD1	RHOA	SPOP	U2AF1
ARID1A	CBFB	CXCR4	ETV4	FLT4	HIST1H3I	KDM5A	MGA	NTRK3	POLE	RICTOR	SPTA1	VEGFA
ARID1B	CBL	CYLD	ETV5	FOXA1	HIST1H3J	KDM5C	MITF	NUP93	PPARG	RIT1	SRC	VHL
ARID2	CCND1	DAXX	ETV6	FOXL2	HIST2H3A	KDM6A	MLH1	NUTM1	PPM1D	RNF43	SRSF2	VTCN1
ARID5B	CCND2	DCUN1D1	EWSR1	FOXO1	HIST2H3C	KDR	MLL	PAK1	PPP2R1A	ROS1	STAG1	WISP3
ASXL1	CCND3	DDR2	EZH2	FOXP1	HIST2H3D	KEAP1	MLL2	PAK3	PPP2R2A	RPS6KA4	STAG2	WT1
ASXL2	CCNE1	DDX41	FAM123B	FRS2	HIST3H3	KEL	MPL	PAK7	PPP6C	RPS6KB1	STAT3	XIAP
ATM	CD274	DHX15	FAM175A	FUBP1	HLA-A	KIF5B	MRE11A	PALB2	PRDM1	RPS6KB2	STAT4	XPO1
ATR	CD276	DICER1	FAM46C	FYN	HLA-B	KIT	MSH2	PARK2	PREX2	RPTOR	STAT5A	XRCC2
ATR	CD74	DIS3	FANCA	GABRA6	HLA-C	KLF4	MSH3	PARP1	PRKAR1A	RUNX1	STAT5B	YAP1
AURKA	CD79A	DNAJB1	FANCC	GATA1	HNF1A	KLHL6	MSH6	PAX3	PRKCI	RUNX1T1	STK11	YES1
AURKB	CD79B	DNMT1	FANCD2	GATA2	HNRNP	KMT2B	MST1	PAX5	PRKDC	RYBP	STK40	ZBTB2
AXIN1	CDC73	DNMT3A	FANCE	GATA3	HOXB13	KMT2C	MST1R	PAX7	PRSS8	SDHA	SUFU	ZBTB7A
AXIN2	CDH1	DNMT3B	FANCF	GATA4	IGF1	KMT2D	MTOR	PAX8	PTCH1	SDHAF2	SUZ12	ZFH3
AXL	CDK12	DOT1L	FANCG	GATA6	IGF1R	KRAS	MUTYH	PBRM1	PTEN	SDHB	SYK	ZNF217
B2M	CDK4	E2F3	FANCI	GEN1	IGF2	LAMP1	MYB	PDCD1	PTPN11	SDHC	TAF1	ZNF703
BAP1	CDK6	EED	FANCL	GID4	IKBKE	LATS1	MYC	PDCD1LG2	PTPRD	SDHD	TBX3	ZRSR2
BARD1	CDK8	EGFL7	FAS	GLI1	IKZF1	LATS2	MYCL1	PDGFRA	PTPRS	SETBP1	TCEB1	
BBC3	CDKN1A	EGFR	FAT1	GNA11	IL10	LMO1	MYCN	PDGFRB	PTPRT	SETD2	TCF3	
BCL10	CDKN1B	EIF1AX	FBXW7	GNA13	IL7R	LRP1B	MYD88	PDK1	QKI	SF3B1	TCF7L2	
BCL2	CDKN2A	EIF4A2	FGF1	GNAQ	INHA	LYN	MYO1D	PDPK1	RAB35	SH2B3	TERC	
BCL2L1	CDKN2B	EIF4E	FGF8	GNAS	HRAS	LZTR1	NAB2	PGR	RAC1	SH2D1A	TER ^b	
BCL2L11	CDKN2C	EML4	FGF9	GPR124	HSD3B1	MAGI2	NBN	PHF6	RAD21	SHQ1	TET1	
BCL2L2	CEBPA	EP300	FGF10	GPS2	HSP90AA1	MALT1	NCOA3	PHOX2B	RAD50	SLIT2	TET2	
BCL6	CENPA	EPCAM	FGF14	GREM1	ICOSLG	MAP2K1	NCOR1	PIK3C2B	RAD51	SLX4	TFE3	
BCOR	CHD2	EPHA3	FGF19	GRIN2A	ID3	MAP2K2	NEGR1	PIK3C2G	RAD51B	SMAD2	TFRC	
BCORL1	CHD4	EPHA5	FGF2	GRM3	IDH1	MAP2K4	NF1	PIK3C3	RAD51C	SMAD3	TGFBR1	

a. Large rearrangements (exon-level CNVs) detected for BRCA1 and BRCA2.

b. TERT promoter region only covered for variant calling.

CNV calling is available for all genes except: DNAJB1, FANCF, FOXL2, HIST1H3A, HIST1H3C, HIST1H3D, HIST1H3E, HIST1H3F, HIST1H3G, HIST1H3H, HIST1H3I, HIST1H3J, HIST2H3A, HIST2H3C, HIST2H3D, HLA-A, HLA-B, HLA-C, KMT2B, KMT2C, KMT2D, TERC, TERT

Table 3: RNA content in the TruSight Oncology 500
TruSight Oncology 500 High-Throughput panels

<i>ABL1</i>	<i>EGFR</i>	<i>FGFR2</i>	<i>MLL</i>	<i>PAX3</i>
<i>AKT3</i>	<i>EML4</i>	<i>FGFR3</i>	<i>MLLT3</i>	<i>PAX7</i>
<i>ALK</i>	<i>ERBB2</i>	<i>FGFR4</i>	<i>MSH2</i>	<i>PDGFRA</i>
<i>AR</i>	<i>ERG</i>	<i>FLI1</i>	<i>MYC</i>	<i>PDGFRB</i>
<i>AXL</i>	<i>ESR1</i>	<i>FLT1</i>	<i>NOTCH1</i>	<i>PIK3CA</i>
<i>BCL2</i>	<i>ETS1</i>	<i>FLT3</i>	<i>NOTCH2</i>	<i>PPARG</i>
<i>BRAF</i>	<i>ETV1</i>	<i>JAK2</i>	<i>NOTCH3</i>	<i>RAF1</i>
<i>BRCA1</i>	<i>ETV4</i>	<i>KDR</i>	<i>NRG1</i>	<i>RET</i>
<i>BRCA2</i>	<i>ETV5</i>	<i>KIF5B</i>	<i>NTRK1</i>	<i>ROS1</i>
<i>CDK4</i>	<i>EWSR1</i>	<i>KIT</i>	<i>NTRK2</i>	<i>RPS6KB1</i>
<i>CSF1R</i>	<i>FGFR1</i>	<i>MET</i>	<i>NTRK3</i>	<i>TMPRSS2</i>

All genes listed are assessed for known and novel fusions; content shaded in gray is analyzed for splice variants.

Automate for efficiency

TruSight Oncology 500 and TruSight Oncology 500 High-Throughput offer manual and automated options to support scalable library prep. Illumina has partnered with Hamilton and Beckman Coulter Life Sciences, leading liquid-handling manufacturers, to produce fully automated workflows for TruSight Oncology 500 assays that support a range of throughput needs. These automated workflows achieve the same high-quality results produced by manual protocols, while reducing hands-on time by ~50%, enabling labs to save on labor costs and improve efficiency.

Add tags for analytical specificity

During library preparation, unique molecular identifiers (UMIs)¹¹ are added to the gDNA or cDNA fragments. These UMIs enable detection of variants at low variant allele frequency (VAF) while simultaneously suppressing errors, providing high analytical specificity.

Enrich libraries to focus efforts

Library preparation is based on proven hybrid-capture chemistry to purify selected targets from DNA- and RNA-based libraries. Biotinylated probes hybridize to regions of interest, which are pulled down using streptavidin-coated magnetic beads, and then eluted to enrich the library pool. Hybridization-based enrichment is a useful strategy for analyzing specific genetic variants in a given sample and reliably sequencing exomes or large numbers of genes (eg, > 50 genes). It delivers dependable results across a wide range of input types and quantities. Hybrid-capture chemistry offers several advantages over amplicon sequencing, including yielding data with fewer artifacts and dropouts. Additionally, hybrid-capture chemistry is fusion agnostic, enabling detection and characterization of known and novel fusions. Unlike amplicon-based approaches, which require confirmatory tests as false-positives can arise, the hybrid-capture method is highly sensitive and can accurately characterize gene fusions with both known and novel partners.

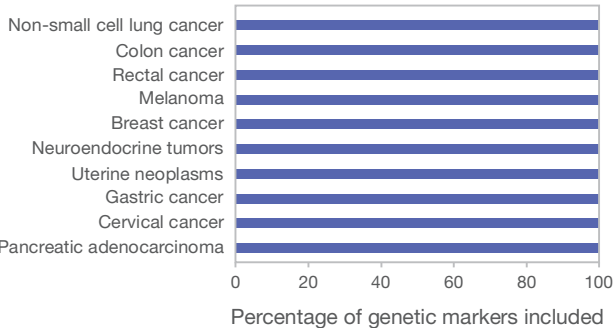


Figure 4: TruSight Oncology 500 content alignment to key guidelines by cancer type—The graph provides examples of content alignment; it is not meant to be all-inclusive.